

Journal of Medicinal Chemistry

© Copyright 1987 by the American Chemical Society

Volume 30, Number 6

June 1987

Communications to the Editor

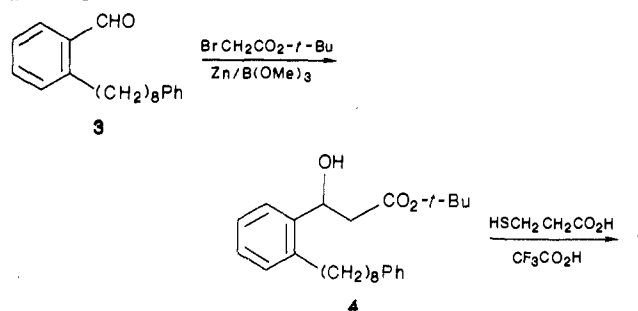
High-Affinity Leukotriene Receptor Antagonists. Synthesis and Pharmacological Characterization of 2-Hydroxy-3-[(2-carboxyethyl)thio]-3-[2-(8-phenyloctyl)phenyl]propanoic Acid

Sir:

The leukotrienes (LTC₄, LTD₄, and LTE₄) have been the focus of intensive research since their identification as the biologically active components of slow reacting substance of anaphylaxis.¹⁻³ Released by sensitized human and animal lung tissue,^{3,4} these natural substances produce prolonged bronchoconstriction,⁵ increased microvascular permeability,^{6,7} and enhanced mucus production.⁸ They have been implicated in a wide variety of immediate hypersensitivity diseases, including asthma, and have also been associated with nonimmunological pulmonary, cardiovascular, and renal diseases.^{9,10}

Most of the pharmacological effects of the leukotrienes appear to be receptor mediated, and [³H]LTD₄-specific binding sites (receptors) have been identified and characterized in human¹¹ and animal tissues.¹²⁻¹⁴ A number of leukotriene antagonists have been reported,¹⁵⁻¹⁷ most

Scheme I



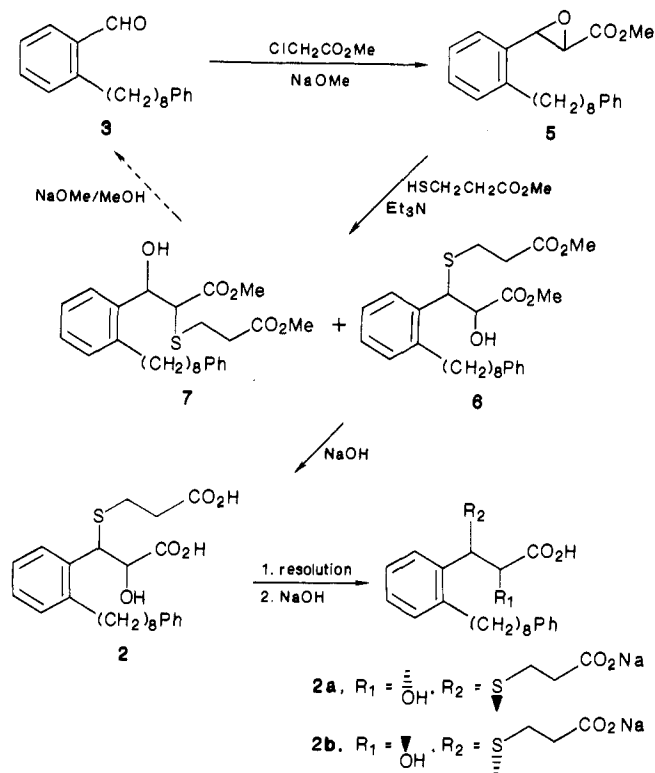
of which bear a close structural similarity with the initial prototype antagonist, FPL 55712.¹⁸ However, these compounds do not exhibit high affinity for LTD₄ receptors ($K_i = 0.5-5 \mu\text{M}$).

Chemical efforts in our laboratories have focused on the design and synthesis of high affinity peptidoleukotriene receptor antagonists. This research has been guided by two key observations. First, 2-norleukotrienes, in which the spacial separation between the eicosanoid carboxyl and thioether functionality of the natural leukotrienes is shortened by one methylene residue, exhibit antagonist properties.¹⁹⁻²¹ Second, the unsaturated triene moiety of LTD₄ may be replaced by a (phenyloctyl)phenyl group, which confers improved chemical and metabolic stability without significant loss of receptor affinity.²² These early studies were not definitive with respect to the importance of the C-5 hydroxyl of LTD₄ for receptor affinity. Deletion of this hydroxyl in antagonist analogues had little effect on potency.^{20,23} In contrast, the agonist potency of LTD₁ is significantly diminished on deletion of the C-5 hydroxyl.²⁴ Further studies in our laboratories have led to the

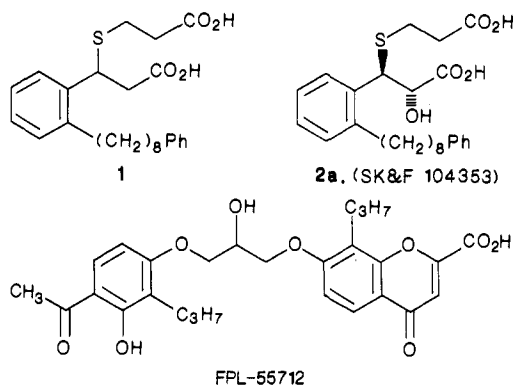
- (1) Murphy, R. C.; Hammarstrom, S.; Samuelsson, B. *Proc. Natl. Acad. Sci. U.S.A.* **1979**, *76*, 4275.
- (2) Morris, H. R.; Taylor, G. W.; Piper, P. J.; Samhoun, M. N.; Tippins, J. R. *Prostaglandins* **1980**, *19*, 185.
- (3) Lewis, R. A.; Austen, K. F.; Drazen, J. M.; Clark, D. A.; Marfat, A.; Corey, E. J. *Proc. Natl. Acad. Sci. U.S.A.* **1983**, *77*, 3710.
- (4) Dahlén, S. E.; Hansson, G.; Hedqvist, P.; Björck, T.; Granstrom, E.; Dahlén, B. *Proc. Natl. Acad. Sci. U.S.A.* **1983**, *80*, 1712.
- (5) Dahlén, S. E.; Hedqvist, P.; Hammarstrom, S.; Samuelsson, B. *Nature (London)* **1980**, *288*, 484.
- (6) Peck, M. J.; Piper, P. J.; Williams, T. J. *Prostaglandins* **1981**, *21*, 315.
- (7) Woodward, D. F.; Weichman, B. M.; Gill, C. A.; Wasserman, M. A. *Prostaglandins* **1983**, *25*, 131.
- (8) Marom, Z.; Shelhamer, J. H.; Bach, M. K.; Morton, D. R.; Kaliner, M. *Am. Rev. Respir. Dis.* **1982**, *126*, 449.
- (9) Lefer, A. M. *Biochem. Pharmacol.* **1986**, *35*, 123.
- (10) Feuerstein, G. J. *Auton. Pharmacol.* **1985**, *5*, 149.
- (11) Lewis, M. A.; Mong, S.; Vessella, R. L.; Croke, S. T. *Biochem. Pharmacol.* **1985**, *34*, 4311.
- (12) Mong, S.; Wu, H. L.; Hogaboom, G. K.; Clark, M. A.; Croke, S. T. *Eur. J. Pharmacol.* **1984**, *102*, 2.
- (13) Pong, S. S.; DeHaven, R. N. *Proc. Natl. Acad. Sci. U.S.A.* **1983**, *80*, 7415.
- (14) Robertson, R. P. *Prostaglandins* **1986**, *31*, 395.
- (15) Fleisch, J. H.; Rinkema, L. E.; Haisch, K. D.; Swanson-Bean, D.; Goodson, T.; Ho, P. P. K.; Marshall, W. S. *J. Pharmacol. Exp. Ther.* **1985**, *233*, 148.
- (16) Young, R. N.; Belanger, P.; Champion, E.; DeHaven, R. N.; Denis, D.; Ford-Hutchinson, A. W.; Fortin, R.; Frenette, R.; Gauthier, J. Y.; Gillard, J.; Guindon, Y.; Jones, T. R.; Kakushima, M.; Masson, P.; Maycock, A.; McFarlane, C. S.; Piechuta, H.; Pong, S. S.; Rokach, J.; Williams, H.; Yoakim, C.; Zamboni, R. *J. Med. Chem.* **1986**, *29*, 1573.

- (17) O'Donnell, M.; Brown, D.; Cohen, N.; Weber, G. F.; Welton, A. F. *Ann. Allergy* **1985**, 278.
- (18) Augstein, J.; Farmer, J. B.; Lee, T. B.; Sheard, P.; Tattersall, M. L. *Nature (London) New Biol.* **1973**, *245*, 215.
- (19) Gleason, J. G.; Ku, T. W.; McCarthy, M. E.; Weichman, B. M.; Holden, D.; Osborn, R. R.; Zabko-Potapovich, B.; Berkowitz, B.; Wasserman, M. A. *Biochem. Biophys. Res. Commun.* **1983**, *117*, 732.
- (20) Perchonock, C. D.; Uzinskas, I.; Ku, T. W.; McCarthy, M. E.; Bondinell, W. E.; Volpe, B. W.; Gleason, J. G.; Weichman, B. M.; Muccitelli, R. M.; DeVan, J. F.; Tucker, S. S.; Vickery, L.; Wasserman, M. A. *Prostaglandins* **1985**, *29*, 75.
- (21) Ku, T. W.; McCarthy, M. E.; Weichman, B. M.; Gleason, J. G. *J. Med. Chem.* **1985**, *28*, 1847.
- (22) Perchonock, C. D.; McCarthy, M. E.; Erhard, K. F.; Gleason, J. G.; Wasserman, M. A.; Muccitelli, R. M.; DeVan, J. F.; Tucker, S. S.; Vickery, L. M.; Kirchner, T.; Weichman, B. M.; Mong, S.; Croke, S. T.; Newton, J. F. *J. Med. Chem.* **1985**, *28*, 1145.
- (23) Gleason, J. G.; Kondrad, K.; Hall, R. F.; Weichman, B. M., unpublished results.

Scheme II

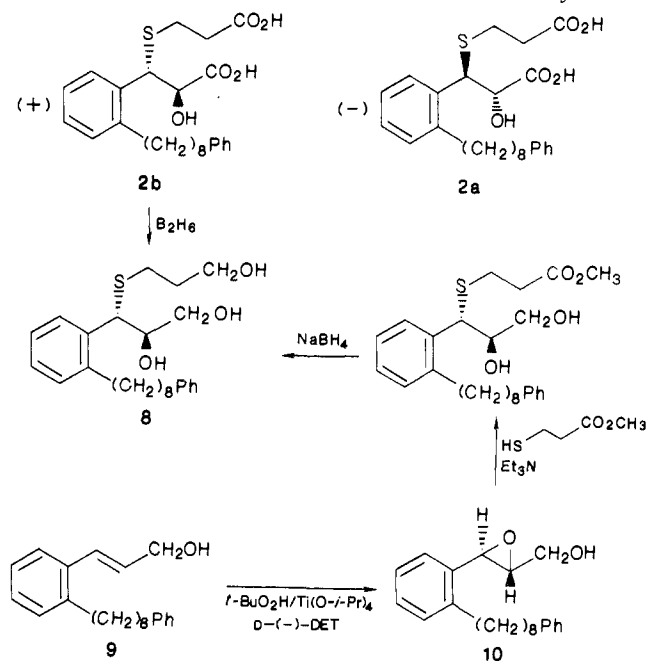


discovery of a series of [(phenyloctyl)phenyl]propionic acids as novel, potent, specific, high-affinity peptidoleukotriene receptor antagonists. The syntheses and pharmacological profile of two members of this class, 3-[(2-carboxyethyl)thio]-3-[2-(8-phenyloctyl)phenyl]propionic acid (1) and 2(*S*)-hydroxy-3(*R*)-[(2-carboxyethyl)thio]-3-[2-(8-phenyloctyl)phenyl]propionic acid (2a, SK&F 104353) are described in this paper.



Chemistry.²⁵ Compound 1 was prepared from (phenyloctyl)benzaldehyde (3)²² as depicted in Scheme I. Trimethyl borate promoted Reformatsky²⁶ reaction of 3 with *tert*-butyl bromoacetate (THF, room temperature, 24 h) afforded the 3-hydroxyarylpropionic ester 4. Displacement of the hydroxyl with mercaptopropionic acid proceeded with simultaneous cleavage of the *tert*-butyl ester (2:1 TFA, CH₂Cl₂, 0 °C, 5 h); conversion to the salt (K₂CO₃, reverse-phase chromatography) afforded the di-

Scheme III. Determination of Absolute Stereochemistry



potassium salt of 1 (mp 270 °C).

The 2-hydroxy analogue 2 was prepared as shown in Scheme II. Darzens condensation of 3 with methyl chloroacetate (1.5 equiv of NaOMe, ether, 0 °C, 2.5 h) afforded the trans-epoxy ester 5 which, on reaction with methyl mercaptopropionate (Et₃N, MeOH, room temperature, 18 h), gave a 1:1 mixture of regioisomers 6 and 7. Treatment of the mixture of regioisomers with methoxide effected a retro-aldol degradation of the undesired isomer 7, affording 6 and recovered aldehyde 3. Ester hydrolysis of 6 (NaOH, MeOH, room temperature) afforded racemic 2, which was resolved as the α -methyl-4-bromobenzylamine salt to give, after conversion to their respective disodium salts (NaOH, MeOH), the enantiomers 2a ($[\alpha]_D^{24}$ -47.0° (c 1, H₂O)) and 2b ($[\alpha]_D^{24}$ +46.6°).

Determination of absolute configuration (Scheme III) was carried out on the dextrorotatory isomer 2b. Thus, diborane reduction of 2b afforded the triol 8 which was identical (NMR, IR, $[\alpha]_D$) to the 2(*R*),3(*S*)-triol prepared by the following independent chiral synthesis.²⁷ Sharpless chiral epoxidation²⁸ of 2-(phenyloctyl)cinnamyl alcohol 9 (*t*-BuO₂H/Ti(*i*-OPr)₄/D-(-)-diethyl tartrate, CH₂Cl₂) afforded the 2(*R*),3(*R*)-epoxide 10, which on reaction with mercaptide and subsequent ester reduction (NaBH₄, THF/H₂O) provided the necessary triol 8.

Pharmacology. Compounds 1, 2a, and 2b were evaluated pharmacologically *in vitro* and *in vivo* and compared to the standard antagonist FPL 55712.¹⁸ Compound 1 competed for specific [³H]LTD₄ binding sites on guinea pig lung membranes with a K_i of 60 ± 8 nM. Introduction of a hydroxyl group adjacent to the carboxylate function significantly enhanced receptor affinity. Thus, 2a, which possesses absolute stereochemistry identical with the natural agonist LTD₄, exhibited high affinity for the receptor (K_i = 5 ± 2 nM), while the enantiomer 2b exhibited significantly reduced affinity (K_i = 180 ± 24 nM).²⁹ These

(24) Lewis, R. A.; Drazen, J. M.; Austen, K. F.; Toda, M.; Brion, F.; Marfat, A.; Corey, E. J. *Proc. Natl. Acad. Sci. U.S.A.* 1981, 78, 4579.

(25) Satisfactory elemental analyses were obtained for all new compounds. All synthetic intermediates were characterized spectroscopically.

(26) Rathke, M. W.; Lindert, A. *J. Org. Chem.* 1970, 35, 3966.

(27) The assignment of the absolute configuration of 2a and 2b is based on the well-documented Sharpless asymmetric epoxidation of allylic alcohols.²⁸

(28) (a) Katsuki, T.; Sharpless, K. B. *J. Am. Chem. Soc.* 1980, 102, 5974. (b) Sharpless, K. B.; Behrens, C. H.; Katsuki, T.; Lee, A. W. M.; Martin, V. S.; Takatani, M.; Viti, S. M.; Walker, F. J.; Woodard, S. S. *Pure Appl. Chem.* 1983, 55, 589.

Table I. In Vitro Antagonist Potency

compd	pK_b^a			
	guinea pig trachea		human bronchus	
	vs. LTD ₄	vs. LTC ₄ ^b	vs. LTD ₄	vs. LTC ₄ ^b
1	7.0	5.1	c	c
2a	8.6	<5.4	8.2	8.3
2b	6.1	c	c	c
FPL 55712	6.7	4.9	5.8	6.4

^a pK_b values for all compounds were calculated at 10 μ M except for 2a (3 μ M). ^b Experiments conducted in the presence of 45 mM L-serine borate complex to inhibit the conversion of LTC₄ to LTD₄. ^c Not performed.

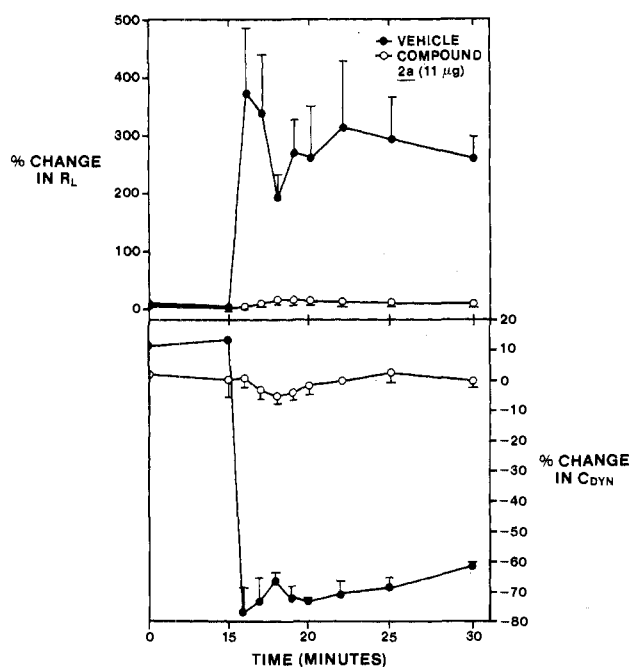


Figure 1. Inhibitory effect of aerosolized 2a on bronchoconstriction induced by aerosolized LTD₄ in the anesthetized guinea pig; (●) control response to LTD₄ (10 ng/animal); (○) 2a (11 μ g/animal) administered 15 min prior to LTD₄ challenge (10 ng).

results are indicative of a highly stereospecific interaction of the antagonist with the receptor and are similar to those reported previously for LTD₄-receptor interactions.¹² These data contrast with the relatively low affinity ($K_i = 1200$ nM) observed for the standard antagonist FPL 55712. Similar results are also observed for 2a in competing for [³H]LTD₄ human lung membrane binding sites ($K_i = 10 \pm 3$ nM).

(29) The observed receptor antagonist activity of 2b (98% ee) may, in part, result from incomplete resolution of 2.

The ability of these compounds to antagonize LTC₄- and LTD₄-induced contractions of guinea pig and human airway smooth muscle is illustrated in Table I. Compound 2a antagonized LTD₄-induced contractions of the guinea pig trachea in a potent and competitive manner³⁰ with a $pA_2 = 8.6$, but was essentially without effect on LTC₄-induced contractions of this tissue. However, on isolated human bronchial tissue, 2a exhibited potent antagonism of contractions induced by either LTD₄ ($pA_2 = 8.2$, slope = 1.14, $n = 5$) or LTC₄ ($pK_b = 8.3$). Compound 1 was 10-fold less potent than 2a, but was considerably more potent than FPL 55712. The enantiomer 2b was weakly active, consistent with its observed lower affinity for the receptor.²⁹ Neither 1 nor 2a elicited a contraction of either guinea pig tracheal or human bronchial tissue at concentrations as high as 10 μ M. Thus, neither compound appears to exhibit partial agonist properties. Moreover, 2a (10 μ M) had no effect on contractions induced by KCl, histamine, carbachol, PGD₂, or the TxA₂ mimic U-44069.

When administered as an aerosol, 2a (0.03%, 100 breaths, 11 μ g total dose) provided complete protection against the changes in airway resistance (R_L) and dynamic lung compliance (C_{DYN}) induced by aerosolized LTD₄ (0.5 μ g/mL, 5 breaths, 10 ng) in anesthetized guinea pigs (Figure 1).

Thus, 2a is a high-affinity leukotriene receptor antagonist, exhibiting potent in vitro and in vivo inhibition of leukotriene-mediated bronchoconstriction. The deshydroxy analogue 1 is similarly an effective antagonist, although with somewhat diminished potency. These compounds represent a novel class of high-affinity leukotriene receptor antagonists that may prove to be of benefit in the treatment of bronchial asthma and other immediate hypersensitivity diseases.

(30) Competitiveness was supported by the observation of parallel shifts in the LTD₄ dose-response curves over a 100-fold concentration range (0.03–3 μ M), which, by Schild analysis, afforded a $pA_2 = 8.6$ and a slope of 0.98 ($n = 4$).

John G. Gleason,* Ralph F. Hall, Carl D. Perchonock
Karl F. Erhard, James S. Frazee, Thomas W. Ku
Karen Kondrad, Mary E. McCarthy, Seymour Mong
Stanley T. Croke, Gloria Chi-Rosso
Martin A. Wasserman, Theodore J. Torphy
Roseanna M. Muccitelli, Douglas W. Hay
Stephanie S. Tucker, Lynne Vickery-Clark

Departments of Medicinal Chemistry, Molecular
Pharmacology, and Pharmacology
Research & Development Division
Smith Kline and French Laboratories
Philadelphia, Pennsylvania 19101

Received November 2, 1986